

**Lecture 38:**

**Data analysis tips**

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# Learning Objectives

- Review of core concepts from Problem Set 8
  - Analyzing multichannel images – marker and sensor
  - Adding data to a vector within a for loop
- Using debugging functions to inspect a for loop
- Measuring time-series statistics from tracked data

# Problem Set 8

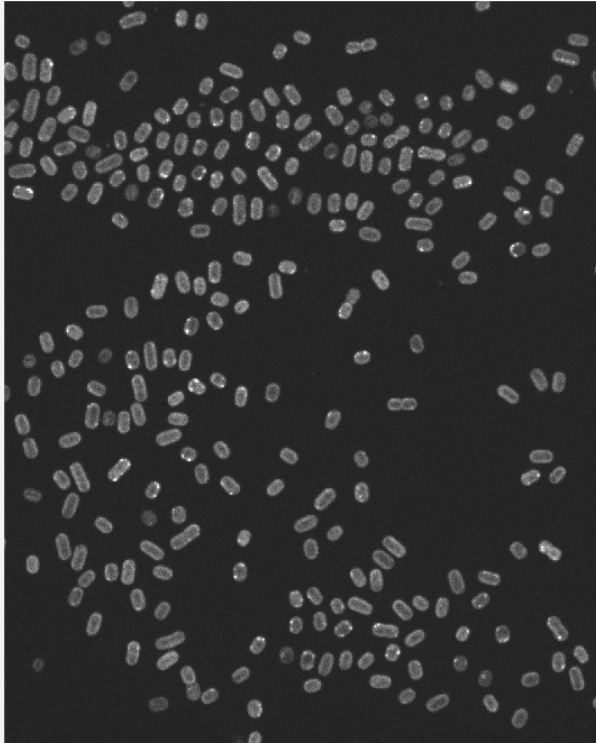
# Question 1

- You were given two images:

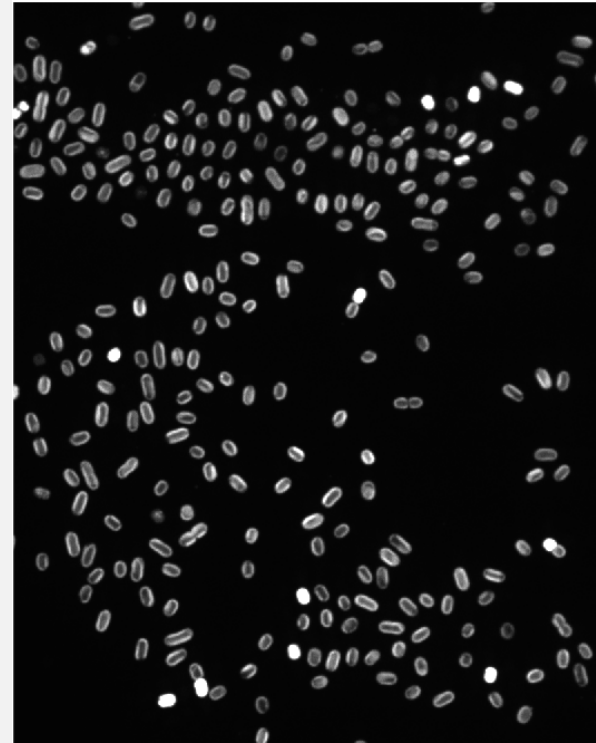
The image `ps8_cyanobacteria_chl.tif` shows the fluorescence arising from natural fluorescence of the chlorophyll pigments, while the image `ps8_cyanobacteria_mNeonGreen.tif` shows the fluorescence from a cell marker that can be used to identify each cell.

# Explanation of the images

mNeonGreen marker



Chlorophyll



**Why can we not use the chlorophyll image to segment cells?**

# Question 1

- The chlorophyll intensity of a cell that is dying will be very bright. Cells that have low chlorophyll intensity are dead.

# Why can we not use the chlorophyll image to segment cells?

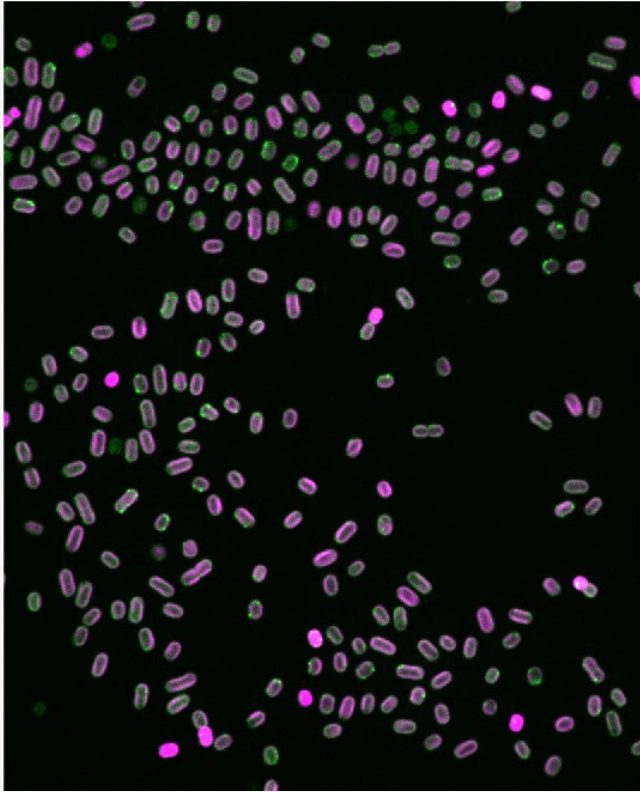
The dead cells will not show up in the chlorophyll channel.



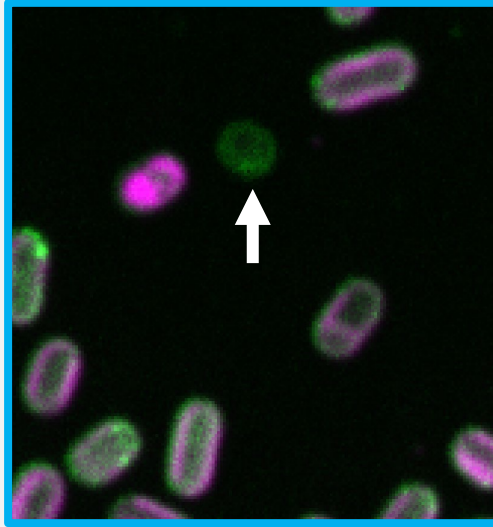
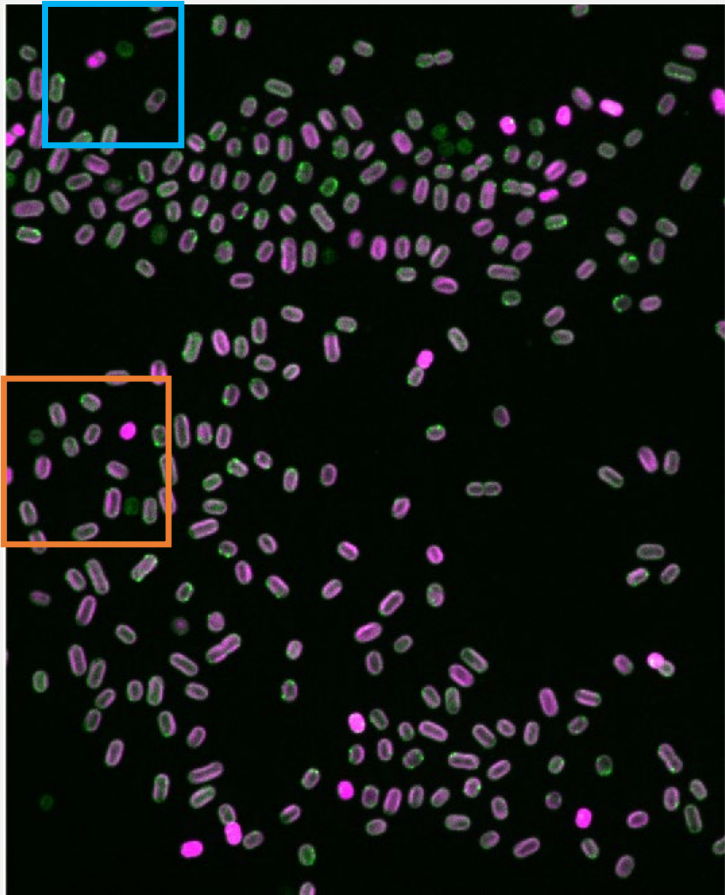
**This is the same concept for any application where you are trying to measure the intensity of a sensor**

You need to have a way of segmenting a cell even when the sensor is dim. Therefore, you cannot segment cells using the sensor image.

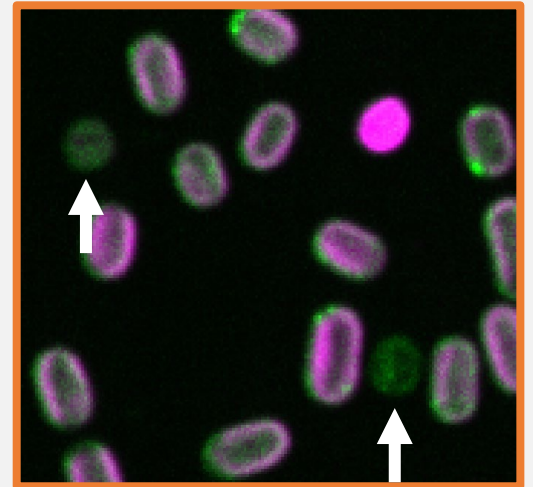
# You can use `imshowpair` to display a "merged" image

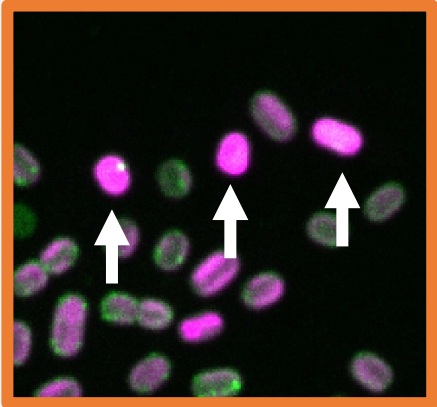
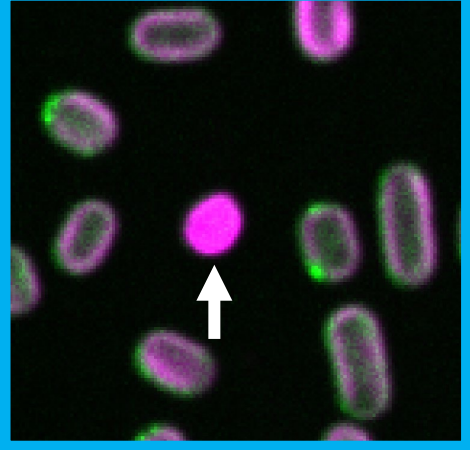
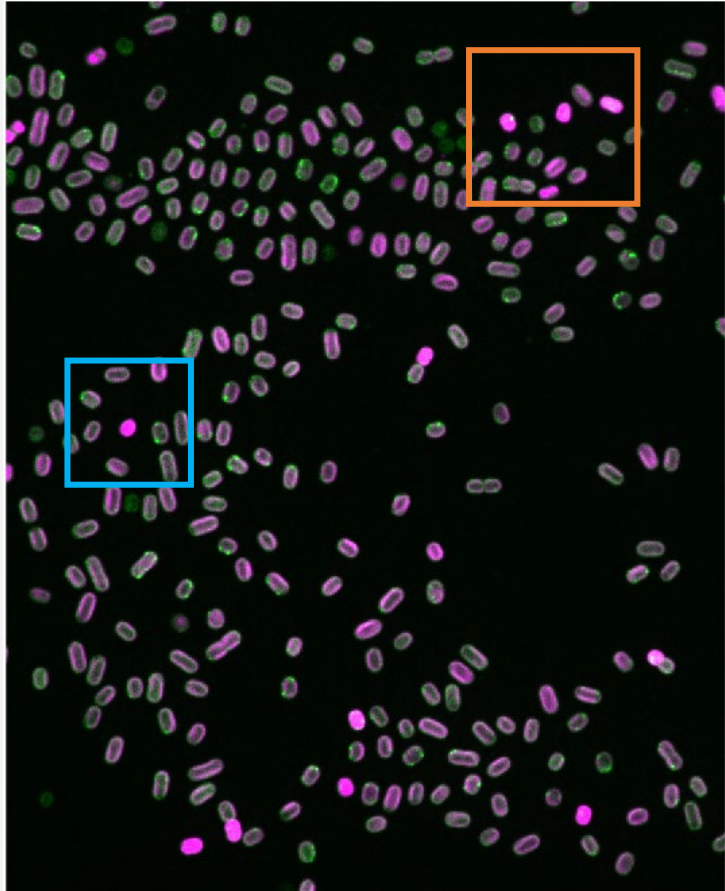


```
Igreen = ...  
imread('ps8_cyanobacteria_mNeonGreen.tif');  
  
Ichl = ...  
imread('imread('ps8_cyanobacteria_mNeonGreen.  
tif');  
  
imshowpair(Igreen, Ichl)
```



These cells are "dead"





These cells are "dying"

# Measuring cell intensities

- The idea is to make a mask using the mNeonGreen image, then use the mask to measure the cell intensities in the chlorophyll channel

# MATLAB Code

```
%Read in images
Igreen = imread('ps8_cyanobacteria_mNeonGreen.tif');
Ichl = imread('ps8_cyanobacteria_chl.tif');

%Make a mask using the mNeonGreen image
mask = Igreen > 700;

celldata = regionprops(mask, Ichl, 'MeanIntensity');
```

# Counting dead (dim) and dying (very bright) cells

# Plotting a histogram

- Syntax:

```
histogram(data, numBins)
```



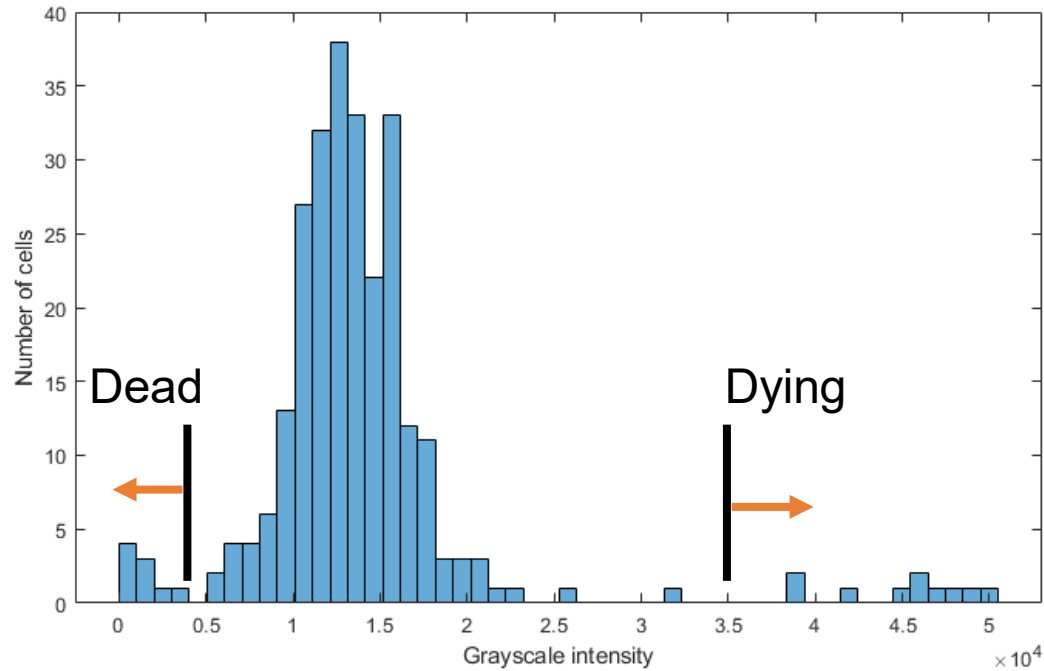
# MATLAB code

```
data = regionprops(mask, Ichl, 'MeanIntensity');
```

```
intensities = cat(1, data.MeanIntensity)
```

```
histogram(intensities, 50)
```

# Plotting a histogram



## Using logical indexing to count the number of cells

```
deadCells = intensities(intensities < 0.5e4);
```

```
numDeadCells = numel(intensities);
```

## Question 2

- In Question 2, you were asked to count the number of cells over time in the time-lapse dataset `PS8_MCF7.tif`
- There are two tips I want to share for analyzing a time-lapse dataset:
  - How to view a timelapse video
  - Storing the number of cells in a vector within the for loop

**Viewing a timelapse video  
(Always good to get an idea of what data to  
expect)**

# Viewing a timelapse video

```
%Get number of frames in the movie
nFrames = numel(imfinfo('ps8_MCF7.tif'));

%Identify cells in each frame of the movie

for iT = 1:nFrames
    I = imread('ps8_MCF7.tif', iT);
    imshow(I, [])
    drawnow
end
```

**Note:** drawnow forces MATLAB to update the figure

# Viewing a timelapse video

- Fiji/ImageJ: <https://imagej.net/software/fiji/>

# Storing data in a vector within a for loop



# Example code

```
nFrames = numel(imfinfo('ps8_MCF7.tif'));
```

```
for iT = 1:nFrames
```

```
    I = imread('ps8_MCF7.tif', iT);
```

```
    mask = imbinarize(I);
```

```
    data = regionprops(mask, 'Area');
```

```
end
```

**Note:** How to calculate the number of cells in the mask?

# Example code

```
nFrames = numel(imfinfo('ps8_MCF7.tif'));
```

```
for iT = 1:nFrames
```

```
    I = imread('ps8_MCF7.tif', iT);
```

```
    mask = imbinarize(I);
```

```
    data = regionprops(mask, 'Area');
```

```
    numCells = numel(data)
```

```
end
```

# Example code

```
nFrames = numel(imfinfo('ps8_MCF7.tif'));
```

```
for iT = 1:nFrames
```

```
    I = imread('ps8_MCF7.tif', iT);
```

```
    mask = imbinarize(I);
```

```
    data = regionprops(mask, 'Area');
```

```
    numCells(iT) = numel(data)
```

```
end
```

**Note:** You can grow a vector by indexing, e.g.:

```
>> A(1) = 1; % A = [1]
```

```
>> A(2) = 10; %A = [1, 10]
```

```
>> A(3) = 23; %A = [1, 10, 23]
```

# Declaring a vector ahead of time

```
nFrames = numel(imfinfo('ps8_MCF7.tif'));
```

```
numCells = zeros(1, nFrames);
```

```
for iT = 1:nFrames
```

```
    I = imread('ps8_MCF7.tif', iT);
```

```
    mask = imbinarize(I);
```

```
    data = regionprops(mask, 'Area');
```

```
    numCells(iT) = numel(data)
```

```
end
```

**Note:** This function creates a row of zeros

# Tips for writing code in a for loop

- Make sure that the code works for one iteration before moving on
- Change the number of iterations to process the first 5 frames
- Set breakpoints to inspect how the code runs line by line within the for loop

# Saving variables in MATLAB

- To save your variables in MATLAB, use the function `save`:

`save(filename)` – saves every variable in the Workspace to a MAT-file

`save(filename, var1, var2, ...)` – saves the variables `var1`, `var2` to a MAT-file

# Examples of using save

- `save('lec38data.mat')`
- `save('lec38data.mat', 'linker')`

# Loading saved data

- Double-click on the .mat file to load the variables back to your workspace
- Alternatively, you can use the function load:

```
load(matFilename)
```

**Note:** When you double-click the mat file, MATLAB will run the load function for you, as shown in the Command Window.



# Analyzing tracked data

**For this section, we will use data in unperturbed.mat. This file contains tracked data from the control dataset.**

# Question

- The sensor in this data becomes bright when cells are not actively dividing. To compare the different datasets, we want to know how the average cell intensity changes over time.

# Clarification of terminology

- Using `regionprops`, we measure the `MeanIntensity` property which is the average intensity of a cell
- Here we are trying to measure the mean `MeanIntensity` of every cell in a frame, for every single frame of the movie
- The challenge is that each track starts and ends at different frames...

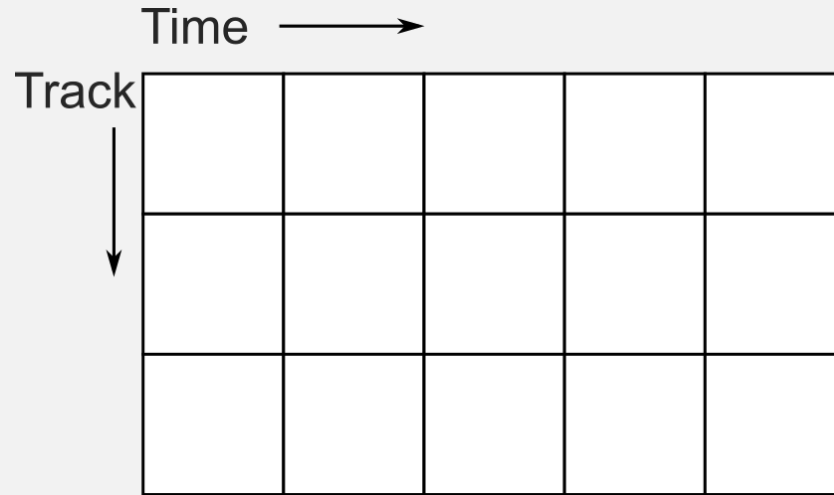
# Track properties

```
>> currTrack = getTrack(linker, 1)
currTrack =
  struct with fields:
```

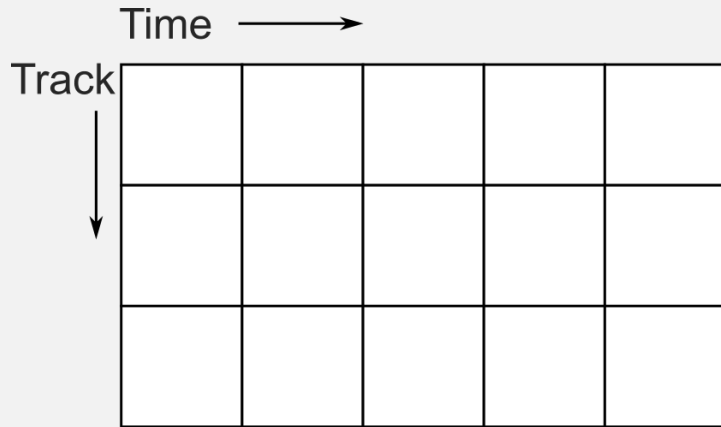
```
        ID: 1
      MotherID: NaN
    DaughterID: NaN
        Frames: [1×30 double]
          Area: [30×1 double]
      Centroid: [30×2 double]
    MeanIntensity: [30×1 double]
```

**Note:** The Frames property tells you when this track existed.

**Basic idea is to collect data into a matrix so we can use the mean function**



# Basic idea is to collect data into a matrix so we can use the mean function



Example:

```
track1.Frames = [1 2 3 4 5];  
track1.MeanIntensity = [10 20 30 40  
50];
```

How to add this data into this matrix?

# Basic idea is to collect data into a matrix so we can use the mean function

Time →

Track ↓

10	20	30	40	50

Example:

```
track1.Frames = [1 2 3 4 5];  
track1.MeanIntensity = [10 20 30 40  
50];
```

How to add this data into this matrix?

```
M(1, track1.Frames) = ...  
    track1.MeanIntensity
```



# Basic idea is to collect data into a matrix so we can use the mean function

Time →

Track ↓

10	20	30	40	50

Example:

```
track2.Frames = [2 3 4 5];
```

```
track2.MeanIntensity = [2 10 8 12];
```

How to add this data into this matrix?

# Basic idea is to collect data into a matrix so we can use the mean function

Time →

Track ↓

10	20	30	40	50
	2	10	8	12

Example:

```
track2.Frames = [2 3 4 5];  
track2.MeanIntensity = [2 10 8 12];
```

How to add this data into this matrix?

```
M(2, track2.Frames) = ...  
    track2.MeanIntensity
```

# Basic idea is to collect data into a matrix so we can use the mean function

Time  $\longrightarrow$

Track  $\downarrow$

10	20	30	40	50
?	2	10	8	12

But... a matrix MUST be rectangular.

So what value should be put in  $M(2, 1)$ ?

# Not-a-Number (NaN)

- NaN stands for Not-a-Number – this is an internal MATLAB designation for a special value that is not a number.
- Essentially any arithmetic operation involving a NaN returns a NaN:

```
>> 10 - NaN
```

```
ans =
```

```
NaN
```

# MATLAB's statistics functions can omit NaNs

```
>> X = [1 2 3; NaN 3 6; 4 6 NaN; 4 7 7];
```

1	2	3
NaN	3	6
4	6	NaN
4	7	7

# MATLAB's statistics functions can omit NaNs

```
>> mean(X, 1, 'omitnan')
```

```
ans =
```

```
3.0000    4.5000    5.3333
```

Mean of elements in first  
column:

$$(1 + 4 + 4)/3 = 3$$

1	2	3
NaN	3	6
4	6	NaN
4	7	7

# Why not use zeros?

```
>> X = [1 2 3; 0 3 6; 4 6 0; 4 7 7]
```

```
X =
```

1	2	3
0	3	6
4	6	0
4	7	7

```
>> mean(X, 1)
```

```
ans =
```

2.2500	4.5000	4.0000
--------	--------	--------

**Note:** A zero would affect the calculation of the mean., e.g. the mean of the first column =  $(1 + 0 + 4 + 4) / 4$

# Declaring a matrix of NaNs

- $M = \text{nan}(\text{numRows}, \text{numCols})$
- Example:  $M = \text{nan}(10, 3)$  generates a matrix of NaNs with 10 rows and 3 columns.



# Example code

- For the example we looked at with the two tracks:

```
%Declare a matrix of NaNs  
allIntensities = nan(2, 5);
```

```
%Index the intensity data using the Frames property to  
%specify the columns  
allIntensities(1, track1.Frames) = track1.MeanIntensity  
allIntensities(2, track2.Frames) = track2.MeanIntensity
```

# Problem Set 10

- The challenge for you to solve in PS10 is to use this method to plot the average intensity of each cell in a frame over the entire movie
- Hint: You should use a for loop