MCDB/BCHM 4312 & 5312 – Quantitative Optical Imaging

Lecture 33:

## Tracking moving objects

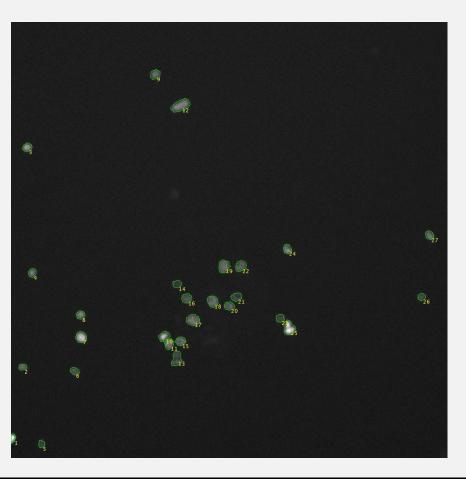
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Date: 12 November 2021



### Learning objectives

- Understand how nearest-neighbor tracking works
- Implement tracking using the linear assignment toolbox



Quantitative microscopy can be used to answer questions such as:

 What happens to cells when treated with X

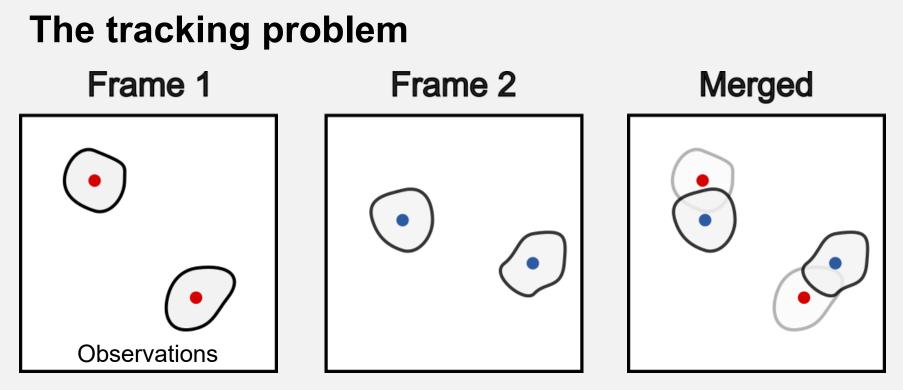
To do this, we need to follow (or <u>track</u>) a cell over time

## The goals of a tracking algorithm

- Identify individual objects in a movie
- Organize this information into a data structure that allows time-series data to be analyzed

## Terminology

- Linking the process of associating objects in one frame with objects in another
- Observations data from objects in current frame (i.e., from regionprops)
- Track a collection of time-series data belonging to a single object (e.g., position, mean intensity over time)

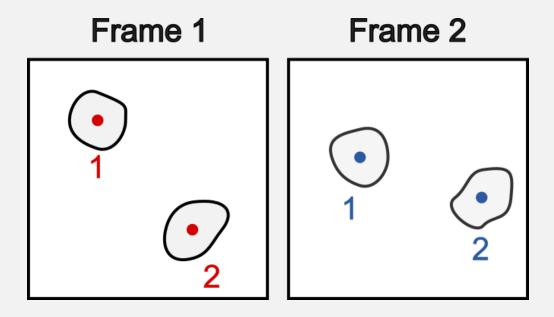


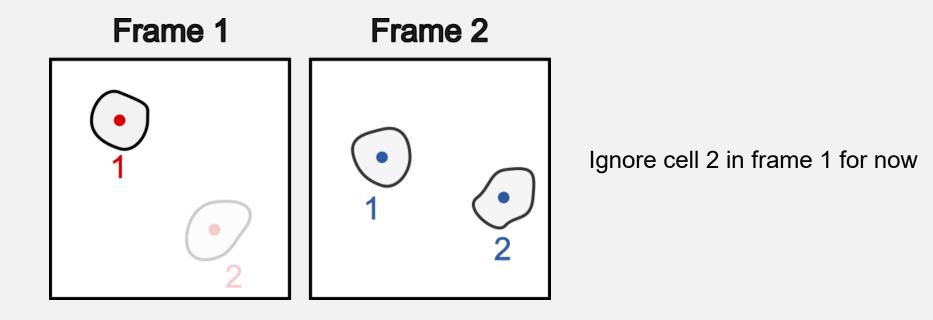
Microscope collects snapshots of the cells. How do we link these objects between frames?

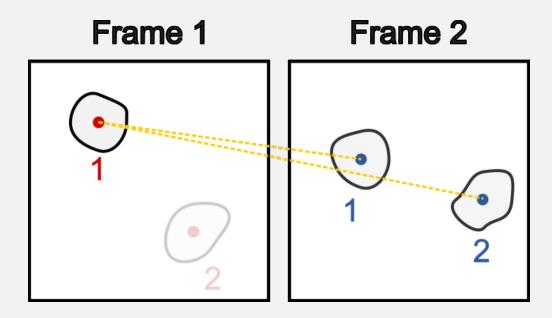
## Nearest neighbor algorithm

Assumption: Cells move slowly compared to frame rate of acquisition

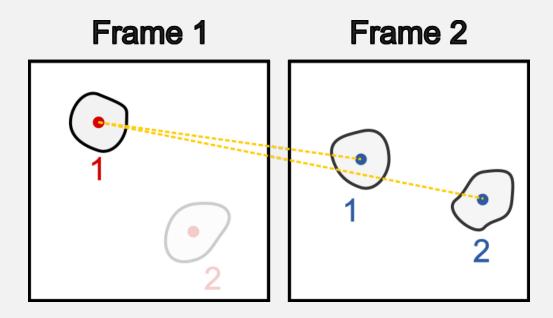
 When comparing object positions between frames, a pair of observations with the shortest distance should be linked together



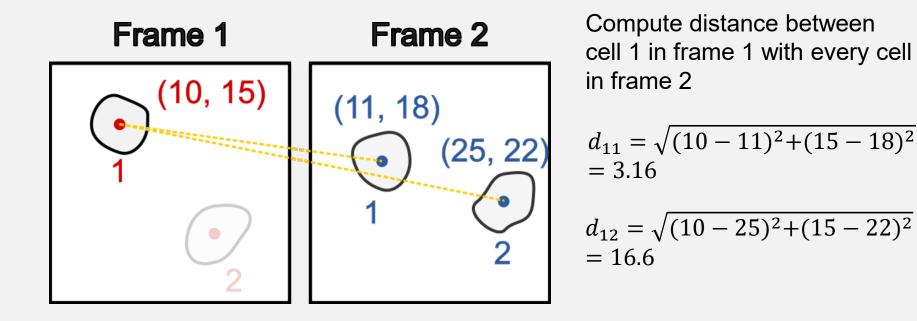


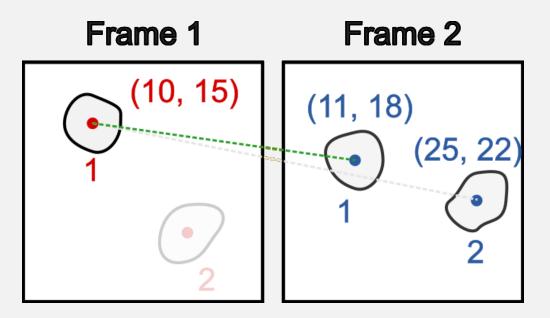


Compute distance between cell 1 in frame 1 with every cell in frame 2



Compute distance between cell 1 in frame 1 with every cell in frame 2



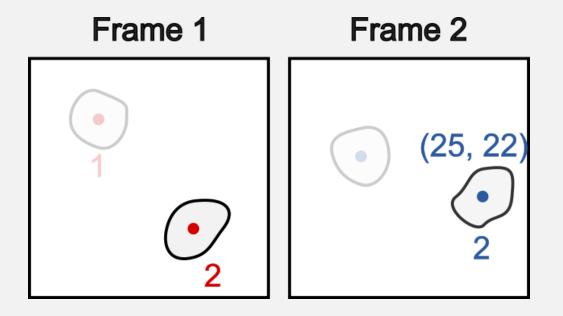


Compute distance between cell 1 in frame 1 with every cell in frame 2

$$d_{11} = \sqrt{(10 - 11)^2 + (15 - 18)^2}$$
  
= 3.16

$$d_{12} = \sqrt{(10 - 25)^2 + (15 - 22)^2}$$
  
= 16.6

Find the shortest distance

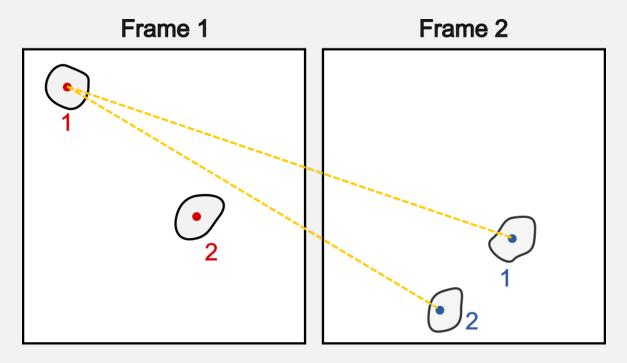


Move to the next cell (cell 2 in frame 1)

But remove linked objects from future consideration

In this example, cell 2 in frame 1 only has a single valid neighbor

#### Prevent "impossible" distances from being linked



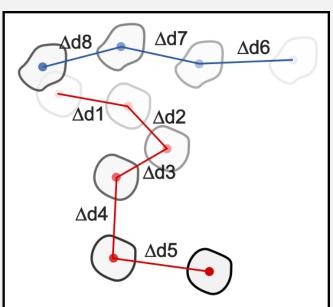
In this example, cell 1 in frame 1 has drifted out of the field of view.

However, if we carry out the nearest-neighbor calculation it would link to cell 1 of Frame 2.

To avoid this, we set a maximum value for this linking distance.

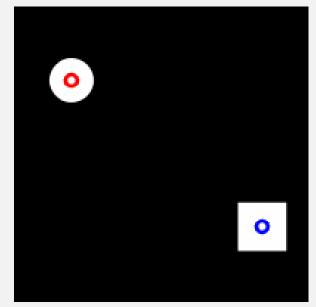
### **Maximum distance for linking**

 The maximum distance for linking should be ~2 – 3x the average step size of objects between frames



#### Issue with the simple nearest-neighbor tracking

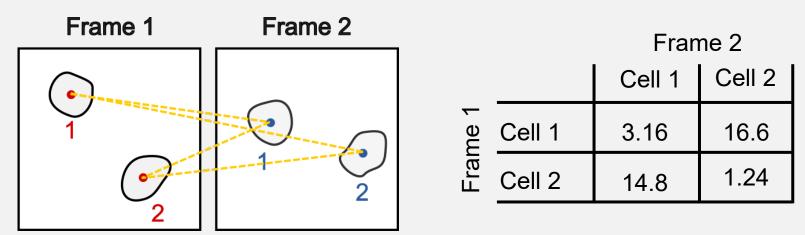
Simple nearest-neighbor makes mistakes when objects "cross paths"



This happens (a lot) when cells are clustered in an image

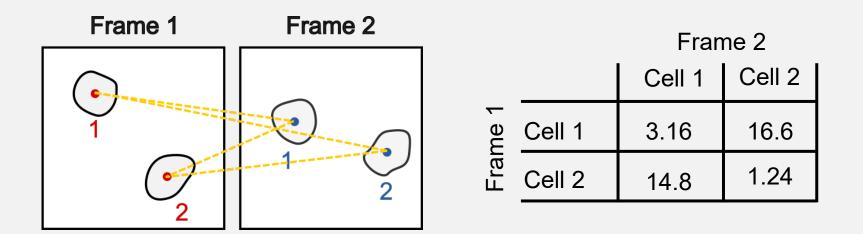
# The linear assignment approach minimizes this problem

 In the linear assignment approach, the distance of every cell in frame 1 to every cell in frame 2 is computed in a matrix

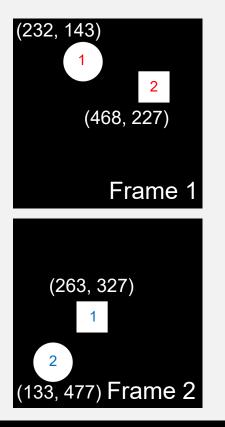


# The linear assignment approach minimizes this problem

- Assignment = choosing a column for each row
- Cells are assigned to <u>minimize the total distance</u>



#### **Nearest-neighbor**



If we used the simple nearest neighbor algorithm, then object 1 in frame 1 (the circle) will have distances

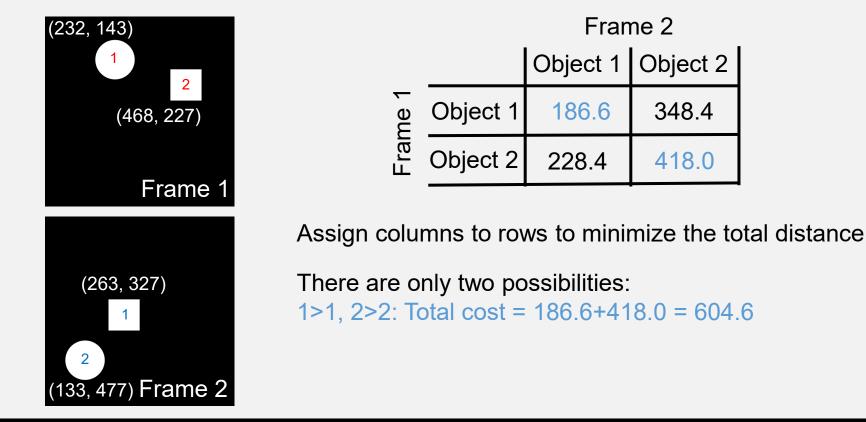
$$d_{11} = \sqrt{(232 - 263)^2 + (143 - 327)^2} = 186.6$$

$$d_{12} = \sqrt{(232 - 133)^2 + (143 - 477)^2} = 348.4$$

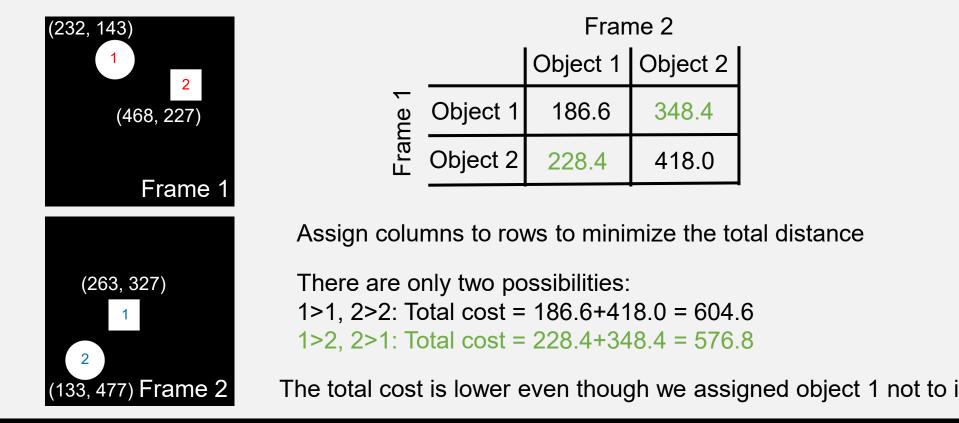
The algorithm would link object 1 (circle) in frame 1 with object 1 (square) in frame 2.

Then object 2 in frame 1 HAS to link with object 1 in frame 2.

## Using the linear assignment approach



## Using the linear assignment approach

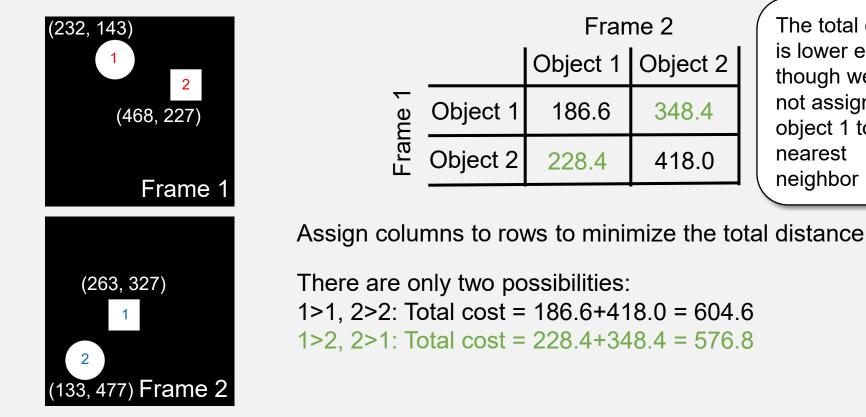


Object 2

348.4

418.0

## Using the linear assignment approach



The total cost is lower even though we did not assign object 1 to its nearest neighbor

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## Summary

- Tracking is an important image analysis problem we want to follow an object over time to get biologically relevant data
- Objects are linked by distance (usually)

 The linear assignment approach links objects by minimizing the total distance of all links

#### You can use other properties to link objects

So far, we've only looked at using distances

 But you can compare sizes, shapes, etc... or even a combination by changing the cost function

• Example: Cost =  $\Delta$ distance +  $\Delta$ area

#### **General limitations**

 Tracking accuracy will decrease depending on cell density – more dense, less accurate

### **Experimental considerations**

- Initial density of cells
  - Assuming cells are growing, how dense can your movie get while still segmenting and tracking cells accurately?

- Frame rate of movie
  - Movie must be recorded fast enough to meet the assumption that cells do not move "much" for tracking
  - But not so frequently that the laser causes photobleaching or phototoxicity

#### **MATLAB** Implementation

### The Linear Assignment toolbox

- This is a general purpose toolbox I wrote to implement the linear assignment tracking algorithm
  - <u>https://github.com/Biofrontiers-ALMC/cell-tracking-toolbox/releases</u>

#### How to use the toolbox

First create a LAPLinker object

#### >> linker = LAPLinker;

## Linking options are in the object properties

 The only option you need to change should be LinkScoreRange

This option specifies the range of valid distances for linking objects: [minDistance, maxDistance]

 By default, the maximum distance is set to 100 px but you should change this after watching the movie

## To change linking score range

Assign the value to a 1x2 matrix:

Example:

#### >> linker.LinkScoreRange = [0 200];

## To track data

- The object was written to use the output struct of regionprops
- To get the code to track objects, you only need to use the method assignToTrack. The syntax is:

linker = assignToTrack(linker, frameNumber, dataStruct)

#### **General code structure**

%Initialize the LAPLinker object

linker = LAPLinker;

```
linker.LinkScoreRange = [0 250];
```

```
for iT = 1:numFrames
```

```
%Read in image
I = imread('file.tif', iT);
```

```
%Make mask, watershed etc...
mask = imbinarize(I);
```

```
%Measure data
data = regionprops(mask, I, 'Centroid', 'MeanIntensity')
```

```
%Track data
linker = assignToTrack(linker, iT, data)
```

#### end

#### Example

- Track cells in the image file nuclearMask.tif
- Using the file L33\_trackingExample.m, <u>add the three lines</u> of code that are missing to track the objects
- Note: This image contains just the mask of cell nuclei. For your homework, you will need to make the masks yourself.

#### Saving tracked data

 Once your code has finished running, you can save data using the function save

## Example: Save the linker object >> save('trackData.mat', 'linker')

To load the data, you can double-click the .mat file to open it in MATLAB

#### Data from the toolbox

To get data, use the method getTrack

Example: To get data from track 1

#### t1 = getTrack(linker, 1)

## Data is stored in a multi-element struct

 Each element of the struct corresponds to an individual object

```
ID: 1
MotherID: NaN
Don't worry about these for now -
these indicate if cells divide
Frames: [1 2 3 4 5 6 7 8 9 10]
Area: [10×1 double]
Centroid: [10×2 double]
```

#### Accessing time-series data

 Data of each cell (measured using regionprops) is stored as a matrix/vector in each named field of the track structure

New time-points are added along the rows

• Example: To get the centroid position of the cell in frame 7
>> position = T1.Centroid(7, :);

#### Accessing time-series data

 The Frames property shows the frames that this object has been tracked in

 Note: The first frame might not be 1 (e.g., if the cell divided, then the first element of Frames will be the first frame the daughter cell was first detected in)

### **Plotting time-series data**

To plot data in a vector, you can use the function plot

plot(xVector, yVector)

#### Example

 This movie was recorded with a frame rate of 10 mins per frame. Plot area vs time in minutes.

#### Example

 This movie was recorded with a frame rate of 10 mins per frame. Plot area vs time in minutes.

```
timeVec = (Frames) * 10;
plot(timeVec, Area)
xlabel('Time (mins)')
ylabel('Area (pixels)')
```

#### Note

- Just like last week, I've supplied an example script if you need help getting started
- The script also includes some lines of code to make a video to show the results of the tracking. You may need to install the Computer Vision Toolbox (Add-Ons > Get Add-Ons and search for this toolbox).
- You don't need to use the code you can write your own from scratch