Computational toolboxes for single cell imaging

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SCR Single Cell Single Molecule Supergroup Feb 17 2020

Microscopy enables individual cells to be observed



Computational tools are needed to analyze the images



- Time-lapse images contain a lot of data (e.g. many cells, intensity, spatial organization, ...)
- Helps avoid tedious analysis by hand as well as experimenter bias (i.e. by drawing expected cell outlines instead of from image data)

Motivation

- Introduce a few MATLAB toolboxes that I have been developing
- Biological context:
 - Studying the carboxysome in cyanobacteria
 - Project with the Cameron lab

The carboxysome plays a central role in carbon fixation within cyanobacteria



- Houses the enzyme Rubisco which catalyzes the primary reactions in carbon fixation
- Important for understanding photosynthesis and improving crop yields

Rubisco is tagged with GFP for visualization

• Rubisco complex • Carbonic anhydrase \bigwedge^{\triangle} Shell proteins



Carboxysome expression is stopped at start of movie



The image analysis pipeline



Detecting cells

- Identified cells using the brightfield image
- Generated mask by intensity thresholding

Brightfield image





Binary mask

Cell clusters are separated by watershedding

Brightfield image



Binary mask



The image analysis pipeline



- Cell length
- Number and position of carboxysomes

Carboxysomes (dots) are identified using the difference of Gaussians filter





- Difference of Gaussians:
 - Blur image with two Gaussian filters with widths σ₁ and σ₂
 - Subtract the blurred images I_{σ2} I_{σ1}
 - The filter keeps structures with size between σ_1 and σ_2

https://micro.magnet.fsu.edu/primer/java/digitalimaging/processing/diffgaussians/index.html

Carboxysome fluorescence

Mask overlay



The image analysis pipeline



- 1. Follow individual cells over time
- 2. Identify cell division

The tracking problem



- Tracking using the linear assignment framework
- Each cell detected in frame 2 is given a score that it is the same cell in frame 1
- The lowest score = the same cell

Jaqaman et al. Nature Methods 5, 695-702 (2008)

Linking score is ratio of overlapping pixels

Overlap score = $\frac{|A \cup B|}{|A \cap B|} = \frac{\text{Number pixels union}}{\text{Number pixels intersect}}$









The linking cost matrix



Overlap score = Number pixels union Number pixels intersect

Frame 2

	А	В	С	D
1				
2				
3				
4				

For each row, assign a column such that total score is minimized

Jonker, R, and Volgenant, A. Computing 38, 325-340 (1987)

The linking cost matrix



Overlap score = Number pixels union Number pixels intersect

Frame 2

	A	В	С	D
1	1.5	Inf	Inf	Inf
2	Inf	4	21	Inf
3	Inf	15	5	Inf
4	Inf	Inf	Inf	1.7

For each row, assign a column such that total score is minimized

Jonker, R, and Volgenant, A. Computing 38, 325-340 (1987)

Cell division identified by overlap score



 Division if two cells in frame 2 overlap the same cell in frame 1

Example of tracking output







Output data structure

- ID unique number assigned to each cell
- MotherID ID of mother cell
- DaughterID ID of daughter cell
- Number of carboxysomes
- Position of carboxysomes
- Cell length

The image analysis pipeline



Output data structure

- ID unique number assigned to each cell
- MotherID ID of mother cell
- DaughterID ID of daughter cell

- Treat data as a binary tree (a common data structure)
- We implemented existing traversal algorithms to analyze these trees

Tree traversal allow us to identify cells with one carboxysome



Cells without carboxysomes do not grow





Quantification of carboxysome activity

Productivity = $\sum_{t} Length(t)$



Productivity = biomass production of a single carboxysome

Clustering different net productivity patterns



Blue: 1 carboxysome Magenta: 0 carboxysomes

Kymograph shows carboxysome location



Inactive carboxysomes change localization and degrade



Summary

- Fluorescence microscopy enables individual cells and structures to be observed and quantified
- Computational tools are necessary for analysis

Available toolboxes

https://biof-git.colorado.edu/biofrontiers-imaging

- BioformatsToolbox Read ND2 (and other microscope) images
 - Uses the Bioformats Java library from openmicroscopy.org
 - Repackaged as a MATLAB class
 - https://biof-git.colorado.edu/biofrontiers-imaging/bioformats-image-toolbox
- LAPLinker Linear Assignment Linking toolbox
 - Object tracking code
 - Lineage/Tree plots
 - https://biof-git.colorado.edu/biofrontiers-imaging/lap-cell-tracker
- CyAn Cyanobacteria Analysis toolbox
 - Segmentation
 - Kymographs
 - Coming soon

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 Working on adding data overlays (e.g. number of spots) to the tree plot

Tracking cells: The linear assignment approach



C = cost of alternative outcome

Alternative outcomes:

- No links cell in frame T was not detected in frame T + 1 (drifted out of FoV or segmentation error)
- New track cell in frame T + 1 was not previously found in frame T (division event or drifted into the FoV or segmentation error)

Example showing image drift between frames



Drift correction



Distance from center to max cross-correlation gives pixel shift

Why microscopy is important in SCSM research: An "ultraproductive" carboxysome?





Carboxysome activity were classified into three distinct phenotypes

